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Determination of pK_a values of fluoroalkanesulfonamides and investigation of their nucleophilicity

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ABSTRACT

The pK_a values of a series of fluoroalkanesulfonylamides were measured by potentiometric titration. Different kinds of alkyl halides and tosylates were employed to investigate the nucleophilicity of fluoroalkanesulfonylamides. Fluoroalkanesulfonylamides with longer fluoroalkyl chain have weaker nucleophilicity.

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1. Introduction

Since the discovery of Prontosil in 1932, a great deal of benzenesulfonylamides have been synthesized and found effective for the treatment of bacterial infections [1]. Common alkanesulfonylamides, however, found little application in the pharmaceutical or agrochemical area at that time [1]. When fluorine was introduced into these common alkanesulfonylamides, their biological activity was satisfactorily improved [1-4]. For example, Trepka and Moore found that fluorinated N-phenylalkanesulfonylamides showed potent herbicidal and anti-inflammatory activities in 1970s [2-4]. Lu recently reported that *N*-alkylfluoroalkanesulfonylamides were environment-friendly insecticides [5]. The introduction of fluorine atoms conferred them high acidity and lipophilicity, resulting in their unique bioactivity. Fluoroalkanesulfonylamides also have special physical and chemical properties. Addition of fluorine atoms to aryl-substituted fluoroalkanesulfonylamides gave a linear acidity increase of 1.47 pK_a units per fluorine and an excellent correlation of partition coefficient in a second-order equation [6]. In the reaction between perfluoroalkanesulfonyla-

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mides and perfluoroalkanesulfonyl fluorides or perfluoroalkanechlorides, bis(perfluoroalkanesulfonyl)imides was sulfonyl successfully synthesized [7]. Perfluoroalkanesulfonylamides reacting with sulfonyl chloride could generate N-sulfinylperfluoroalkanesulfonylamides which were very important intermediates [8,9]. N,N-Dichlorofluoroalkanesulfonylamides, another kind of reactive species, could be conveniently synthesized by the reaction of fluoroalkanesulfonylamides with aqueous KOH and chlorine gas [10]. Transformation of perfluoroalkanesulfonylamides to the corresponding R_fSO₂NHK by solid potassium hydroxide in 1,2dimethoxyethane, the nucleophilicity was increased strongly enough to react with CS₂ and RX forming (S,S)-dialkyl N-(perfluoroalkylsulfonyl)carbodithioimidates [11]. N,N-Dialkylfluoroalkanesulfonylamides were obtained as undesired products with aqueous KOH using DMF as the solvent in the same reaction [11]. However, nature of the reaction was still unknown. To the best of our knowledge, efforts to investigate the details of the nucleophilicity and acidity of fluoroalkanesulfonylamides as well as the compatibility of fluoroalkanesulfonylamides with different kinds of electrophiles have not already been made till now. Therefore, it is important and meaningful to gain more insight into the acidity and nucleophilicity of fluoroalkanesulfonamides, even though some nucleophilic reactions of perfluoroalkanesulfonylamides have been reported [12]. Herein, we report the results.

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2. Results and discussion

Fluoroalkanesulfonylamides could be conveniently synthesized by using fluoroalkanesulfonyl fluorides as starting materials [5,13]. **1a–c** and **1e–g** were favorably obtained in high yield when fluoroalkanesulfonyl fluorides reacted with liquid ammonia at a temperature below –60 °C (entries 1–3 and 5–7, Table 1). Treatment of **1c** with NaOH in methanol, **1d** was satisfactorily formed according to our previous work [14]. FO₂S(CF₂)₂O(CF₂)₂. SO₂F reacted with liquid ammonia, giving **1h** in 97% yield (entry 8). Meanwhile, **1i** was generated from FO₂S(CF₂)₂O(CF₂)₂SO₂F in the same reaction condition (entry 9).

There are several methods for the determination of pK_a value [15]. Among them, potentiometric titration has been the standard method for pK_a measurement due to its accuracy and reproducibility [16]. Furthermore, most fluorinated compounds are often water-insoluble and have unfavorable molar absorption coeffi-

Table 1

Preparation of fluoroalkylsulfonamides and their pK_a values. RfSO₂ + NH₃ (liquid) $\stackrel{<-60 \circ Cacid}{\longrightarrow}$ RfSO₂ NH_{2 1a-i}.

Entry	RfSO ₂ NH ₂	Yield ^a (%)	pK _{a1} ^b	pK _{a2} ^b
1 ^c	CF ₃ SO ₂ NH ₂	80 (1a)	6.28	9.37
2	$Cl(CF_2)_2SO_2NH_2$	88 (1b)	6.17	7.82
3	$I(CF_2)_2O(CF_2)_2SO_2NH_2$	92 (1c)	6.22	7.80
4 ^d	$H(CF_2)_2O(CF_2)_2SO_2NH_2$	87 (1d)	5.83	7.75
5	$I(CF_2)_4O(CF_2)_2SO_2NH_2$	94 (1e)	5.70	7.66
6	$I(CF_2)_6O(CF_2)_2SO_2NH_2$	93 (1f)	4.22	7.57
7	$Cl(CF_2)_6O(CF_2)_2SO_2NH_2$	98 (1g)	5.74	7.56
8 ^c	$H_2NO_2S(CF_2)_2O(CF_2)_2SO_2NH_2$	97 (1h)	5.67	7.92
9	$H_2NO_2S(CF_2)_2O(CF_2)_4O(CF_2)_2SO_2NH_2$	94 (1i)	5.90	7.80

^a Isolated yield.

^b The pK_a values were determined in MeOH using KOH as the base and METTLER TOLEDO T70 3.0.0 as potentiometric titrator.

^c **1a** and **1h** are water soluble.

^d **1d** was prepared from the hydrodehalogenation of **1c** [14].

Table 2	
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Nucleophilicity of the fluoroalkanesulfonylamides.

 $RfSO_2 + RX \xrightarrow{K_2CO_3} RfSO_2NR_2.$

Entry	$RfSO_2NH_2$	RX ^a	RfSO ₂ NR ₂	Conditions (temperature, time)	Yield ^b (%)
1 ^c	1a	CH ₃ I	$CF_3SO_2N(CH_3)_2$	Reflux, 14 h	100 (2aa)
2	1a	C_2H_5I	$CF_3SO_2N(C_2H_5)_2$	Reflux, 22 h	90 (2ab)
3 ^c	1a	n-C ₃ H ₇ Br	$CF_3SO_2NH(n-C_3H_7)$	90°C, 2 d	33 (2ac)
			$CF_3SO_2N(n-C_3H_7)_2$		57 (2ad)
4 ^c	1a	n-C ₄ H ₉ I	$CF_3SO_2NH(n-C_4H_9)$	90 °C, 2 d	15 (2ae)
			$CF_3SO_2N(n-C_4H_9)_2$		85 (2af)
5 ^c	1a	n-C ₄ H ₉ Br	$CF_3SO_2NH(n-C_4H_9)$	90°C, 2 d	28 (2ae)
			$CF_3SO_2N(n-C_4H_9)_2$		66 (2af)
6 ^c	1a	n-C ₄ H ₉ Cl	-	90 °C, 2 d	-
7 ^d	1a	PhCH ₂ Br	CF ₃ SO ₂ NHCH ₂ C ₆ H ₅	Reflux, 22 h	6 (2ag)
			$CF_3SO_2N(CH_2C_6H_5)_2$		29 (2ah)
8	1a	PhCH ₂ Br	$CF_3SO_2N(CH_2C_6H_5)_2$	Reflux, 22 h	86 (2ah)
9	1a	Ts(OCH ₂ CH ₂) ₂ OH	$CF_3SO_2N[(CH_2CH_2O)_2H]_2$	Reflux, 2 d	71 (2ai)
10	1a	Ts(OCH ₂ CH ₂) ₃ OH	$CF_3SO_2N[(CH_2CH_2O)_3H]_2$	Reflux, 2 d	68 (2aj)
11 ^c	1a	Ts(OCH ₂ CH ₂) ₂ OTs		Reflux, 21 h	75 (2ak)
			CF ₃ SO ₂ N_O		
12	1a	Ts(OCH ₂ CH ₂) ₃ OTs	^-^	90 °C, 6d	9 (2al)
			F ₃ Co ₂ S ^{-N} , O		
13	1d	CH ₃ I	$H(CF_2)_2O(CF_2)_2SO_2N(CH_3)_2$	Reflux, 19h	88 (2da)
14	1d	C ₂ H ₅ I	$H(CF_2)_2O(CF_2)_2SO_2N(C_2H_5)_2$	Reflux, 2 d	89 (2db)
15	1d	n-C ₃ H ₇ Br	$H(CF_2)_2O(CF_2)_2SO_2NH(n-C_3H_7)$	90°C, 3 d	37 (2dc)
		5.7	$H(CF_2)_2O(CF_2)_2SO_2N(n-C_3H_7)_2$		41 (2dd)
16	1d	n-C ₄ H ₉ I	$H(CF_2)_2O(CF_2)_2SO_2NH(n-C_4H_9)$	90°C, 2 d	43 (2de)
			$H(CF_2)_2O(CF_2)_2SO_2N(n-C_4H_9)_2$		45 (2df)
17	1d	PhCH ₂ Br	$H(CF_2)_2O(CF_2)_2SO_2N(CH_2C_6H_5)_2$	Reflux, 17 h	88 (2dg)
18	1d	CH ₂ =CHCH ₂ Br	$H(CF_2)_2O(CF_2)_2SO_2N(CH_2CH=CH_2)_2$	Reflux, 31 h	75 (2dh)
19 ^c	1d	Ts(OCH ₂ CH ₂) ₂ OTs		Reflux, 2 d	87 (2di)
		(2 2/2	H(CF ₂) ₂ O(CF ₂) ₂ SO ₂ N		
20	1h	CH₃I	$(CH_3)_2NO_2S(CF_2)_2O(CF_2)_2SO_2N(CH_3)_2$	Reflux, 18 h	35 (2ha)
21	1h	C ₂ H ₅ I	$(C_2H_5)_2NO_2S(CF_2)_2O(CF_2)_2SO_2N(C_2H_5)_2$	90 °C, 31 h	26 (2hb)
22	1h	CH ₂ =CHCH ₂ Br	(CH ₂ =CHCH ₂) ₂ NO ₂ S(CF ₂) ₂ O(CF ₂) ₂ SO ₂ N(CH ₂ CH=CH ₂) ₂	90 °C, 31 h	31 (2hc)
23	1h	PhCH ₂ Br	(C ₆ H ₅ CH ₂) ₂ NO ₂ S(CF ₂) ₂ O(CF ₂) ₂ SO ₂ N(CH ₂ C ₆ H ₅) ₂	Reflux, 3 d	28 (2hd)
24	1i	CH₃I	$(CH_3)_2NO_2S[(CF_2)_2O(CF_2)_2]_2SO_2N(CH_3)_2$	Reflux, 18 h	30 (2ia)
25	1i	C ₂ H ₅ I	$(C_2H_5)_2NO_2S[(CF_2)_2O(CF_2)_2]_2SO_2N(C_2H_5)_2$	90 °C, 33 h	27 (2ib)
26	1i	CH ₂ =CHCH ₂ Br	$(CH_2=CHCH_2)_2NO_2S[(CF_2)_2O(CF_2)_2]_2SO_2N(CH_2CH=CH_2)_2$	90 °C, 31 h	31 (2ic)
27	1i	PhCH ₂ Br	$(C_6H_5CH_2)_2NO_2S[(CF_2)_2O(CF_2)_2]_2SO_2N(CH_2C_6H_5)_2$	Reflux, 2 d	86 (2id)

^b Isolated yield.

 $^{\rm c}\,$ The yields of the products were determined by $^{19}{\rm F}\,{\rm NMR}.$

^d The molar ratio of **1a** to PhCH₂Br was 1:1.1.

cients [17]. Therefore, potentiometric titration was employed for the pK_a determination of **1a-1i**. The results are shown in Table 1. In the cases of 1c, 1e and 1f, the titration must be conducted in darkness to avoid the hydrodehalogenation [14]. Elongation of the fluoroalkyl chain led to a gentle decrease of pK_{a1} and pK_{a2} (entries 1 and 2, entries 3, 5 and 6). The pK_{a1} difference (1.48) between **1e** and 1f was larger than that (0.52) between 1c and 1e (entries 3, 5 and 6). It seems that the halogen bonding interaction between iodine and negative ion of nitrogen in RfNHK (1f) played important role in stabilizing the anion, which apparently decreased the pK_a value [18]. Replacement of iodine with hydrogen in 1c resulted in increase of the acidity (1d), which was very close to 1e (entries 4 and 5). This was indicated that the hydrogen bonding interaction between the hydrogen atom and the negative ion greatly stabilized the anion, which further decreased the pK_a value of product. Substitution of the terminal iodine (1f) with chlorine (1g) increased the pK_{a1} value by 1.52. But little difference was found in their pK_{a2} values (entries 6 and 7). Comparing the pK_a value of **1h** and 1i with 1d and 1e, no significant difference was observed in either their pK_{a1} or pK_{a2} values (entries 8 and 9), indicating that the introduction of another sulfonylamide group into fluoroalkanesulfonylamides had little influence on their acidity.

Generally, the pK_a values of fluoroalkanesulfonamides are lower than the non-fluorinated analogues, thus leading to their weaker nucleophilicity. Although the *N*-substituted fluoroalkanesulfonylamides could be synthesized from the reaction of fluoroalkanesulfonyl fluorides and the corresponding substituted amines, most of the amines are not commercially available, thus making the procedure not much versatile. Therefore, it is interesting to investigate the nucleophilic substitution of fluoroalkanesulfonylamides.

As shown in Table 2, the nucleophilic reactions of fluoroalkanesulfonylamides were much influenced by the electrophilicity of the electrophiles. Strong electrophilic alkyl halides favored the alkylation reaction. For example, treatment of 1a with methyl iodide and K₂CO₃ in refluxing THF for 14 h gave the dimethylated product, 2aa, in 100% yield (entry 1). Replacing methyl iodide with ethyl iodide under the same condition, however, longer reaction time was necessary for complete conversion (entry 2). Taking n- $C_{3}H_{7}Br$ or $n-C_{4}H_{9}I$ instead of ethyl iodide as the electrophile, the reaction became more difficult and time consuming (entries 3 and 4). Higher temperature was needed for completion of the reaction. Mono- and di-substitution occurred simultaneously while increasing the temperature to 90 °C. Similar results were obtained in the reaction between **1a** and n-C₄H₉Br (entry 5). No reaction happened in the case of n-C₄H₉Cl (entry 6). The electrophilicity of the alkyl halides also has influence on the ratio of the products. Lower yield of di-substituted product **2af** was formed when n-C₄H₉Br was used as compared with n-C₄H₉I (entries 4 and 5). Nevertheless, the formation of the di-substituted fluoroalkanesulfonvlamides was still dominant. The di-substitution was accelerated by the first alkylation. It was proposed that the electron donating effect of the first alkyl group on the N atom improved its nucleophilicity, thus resulting in the favorable formation of di-substituted ones. In addition, the yield was obviously affected by the reactant ratio. The dibenzylated product 2ah was obtained in 86% yield when 2.2 equivalent of PhCH₂Br was used. However, only 6% yield of monobenzylated product 2ag and 29% yield of 2ah were formed when 1.1 equivalent of PhCH₂Br was employed under the same condition (entries 7 and 8). Di-substituted fluoroalkanesulfonylamide was also the main product even in the presence of insufficient of benzyl bromide (entry 8).

Similar results were obtained when tosylates were used as electrophiles (entries 9 and 10). It is worth noting that intramolecular cyclization happened preferentially in the reaction of **1a** with di(ethylene glycol) di-*p*-tosylate, even 2.2 equivalent of

tosylate $T_s(OCH_2CH_2)_2OTs$ was used (entry 11). While in the case of tri(ethylene glycol) di-*p*-tosylate, $T_s(OCH_2CH_2)_3OTs$, only 9% yield of intermolecular cyclization product **2al** was formed (entry 12). It seems that the chelating interaction between potassium cations and azacrown ether was inhibited by the strong electron withdrawing effect of trifluoromethanesulfonyl group on N atoms, thus leading to no template effects of potassium cation on the ring closing procedure [19].

The length of fluoroalkyl chain not only affects the pK_a value of the fluoroalkanesulfonamides as aforementioned but also influences their nucleophilicity. As compared with trifluoromethanesulfonamide **1a**, longer reaction time was required for 1,1,2,2-tetrafluoro-2-(1,1,2,2-tetrafluoroethoxy)ethanesulfonamide **1d** to achieve the similar results (entries 13–16). Elongation of the fluoroalkyl chain increased the electron withdrawing effect of the fluoroalkylsulfonyl group, leading to the lower nucleophilicity of **1d**.

In the case of di-sulfonamides **1h** and **1i**, tetraalkylated products were readily obtained (entries 20–27). But the yields were slightly low. Increasing the reaction temperature or prolonging the reaction time had little improvement of the transformation.

3. Conclusion

In summary, the pK_{a1} and pK_{a2} values of a series of fluoroalkanesulfonylamides were measured by potentiometric titration. The sulfonylamides with longer fluoroalkyl chain showed stronger acidity. The terminal substituents also have influences on their pK_a values. Different alkyl halides and tosylates were employed to investigate the nucleophilicity of fluoroalkanesulfonylamides. It was demonstrated that elongation of the fluoroalkyl chain reduced the nucleophilicity of the sulfonamides, which is, to some extent, in accordance with the decrease of their pK_a values. Further investigation on the applications of these fluoroalkanesulfonylamides is going on.

4. Experimental

4.1. General

Unless otherwise stated, NMR spectra were recorded in deuterated chloroform at 300 MHz (¹H NMR) and 282 MHz (¹⁹F NMR). ¹³C NMR spectra were recorded at 75 or 100 MHz in CDCl₃. All chemical shifts were reported in ppm relative to TMS and CFCl₃ (positive for downfield shifts) as external standards. Melting points were measured and uncorrected. MeOH was distilled from 4A molecular sieves before use. Analytical pure THF was used without further purification. All fluoroalkanesulfonyl fluorides except $FO_2S(CF_2)_2O(C Cl(CF_2)_2SO_2F_1$ $FO_{2}S(CF_{2})_{2}O(CF_{2})_{2}SO_{2}F$ and $F_2_4O(CF_2)_2SO_2F$ were obtained from commercial source. $Cl(CF_2)_2SO_2F_1$ $FO_2S(CF_2)_2O(CF_2)_2SO_2F$ and $FO_2S(CF_2)_2O(C F_2_4O(CF_2)_2SO_2F$ were prepared according to the literature [13c,20]. 1d was prepared according to our previous work [14]. The pK_{a1} and pK_{a2} values of fluoroalkanesulfonylamides were measured in MeOH. Samples in the range from $5\times 10^{-5}\,\text{M}$ to $1\times 10^{-2}\,M$ were dissolved in methanol and titrated with KOH (0.02 M in the same solvent). Potentiometric titrator (METTLER TOLEDO T70 3.0.0) was employed to monitor the whole titration process.

4.2. Typical procedure for the preparation of 1a

In a 500 mL three-neck flask equipped with katathermometer and magnetic stirrer, CF_3SO_2F (170 g, 1.12 mol) was bubbled into liquid NH₃ (300 mL) within 1.5 h (during this procedure, temperature of the reaction system inevitably rose up to about -60 °C. But it returned to -78 °C quickly when bubbling stopped. It was the same case for **1b** and **c** and **1e** and **i**). The reaction was stirred at -78 °C for 2 h under nitrogen atmosphere. Then the excess of ammonia was removed by evaporation. The residue was added in dioxane (500 mL), acidified with anhydrous HCl till the PH value is below 2 and filtered. The filtrate was then distilled to give a light yellow powder. Sublimation of the crude powder under vacuum gave a white crystal **1a** (134 g, 80%). ¹H NMR (CD₃COCD₃): δ 7.60 (s, 2H). ¹⁹F NMR (CD₃COCD₃): δ 80.4 (s, 3F).

4.3. Typical procedure for the preparation of 1b and c, 1e-g and 1i

1,1,2,2-Tetrafluoro-2-(1,1,2,2-tetrafluoro-2-iodoethoxy)ethanesulfonyl fluoride (9.80 g, 23.0 mmol) was added dropwise into liquid NH₃ (45 mL) which was cooled by acetone/dry ice bath. After being stirred at -78 °C for 2 h, the excess of ammonia was removed by evaporation. The resulting white slurry was acidified to a pH of 2 with diluted H₂SO₄ and extracted with diethyl ether (80 mL), washed with water (3 × 20 mL), dried over anhydrous Na₂SO₄, concentrated under reduced pressure and dried in vacuum to give the crude product. 8.95 g of pure **1c** (21.2 mmol, 92%) was obtained as a white solid by column chromatography on silica gel using dichloromethane as the eluent. ¹H NMR: δ 5.36 (s, 2H). ¹⁹F NMR: δ –115.8 (s, 2F), –84.8 (m, 2F), –81.0 (t, *J* = 12.4 Hz, 2F), –64.3 (t, *J* = 12.4 Hz, 2F).

1b: white solid; mp 76–77 °C. Yield: 88%. ¹H NMR: δ 5.50 (s, 2H). ¹⁹F NMR: δ –110.7 (s, 2F), –65.6 (s, 2F). MS (ESI, *m/z*, %): 214.0 [M]⁻ (100). IR (KBr): 3377, 3284, 1539, 1361, 1198, 1180, 1162, 1131, 1085, 833, 698, 614, 588, 540, 493, 428 cm⁻¹. Anal. Calcd. for C₂H₂ClF₄NO₂S: C, 11.14; H, 0.94; N, 6.50. Found: C, 11.15; H, 0.93; N, 6.54.

1e: white solid. Yield: 94%. ¹H NMR: δ 5.32 (s, 2H). ¹⁹F NMR: δ -124.8 (m, 2F), -116.7 (d, *J* = 7.9 Hz, 2F), -113.8 (m, 2F), -83.5 (m, 2F), -81.5 (t, *J* = 12.9 Hz, 2F), -59.8 (t, *J* = 13.9 Hz, 2F).

1f: white solid; mp 90–92 °C. Yield: 93%. ¹H NMR: δ 5.40 (s, 2H). ¹⁹F NMR: δ –124.6 (s, 2F), –121.5 (s, 2F), –120.5 (s, 2F), –115.8 (s, 2F), –112.5 (s, 2F), –82.2 (m, 2F), –80.6 (t, *J* = 13.4 Hz, 2F), –58.6 (t, *J* = 14.5 Hz, 2F). MS (ESI, *m/z*, %): 621.8 [M]⁻ (100). IR (KBr): 3607, 3374, 3189, 1563, 1388, 1369, 1328, 1209, 1144, 1090, 1038, 999, 942, 842, 692, 647, 622, 543, 491 cm⁻¹. Anal. Calcd. for C₈H₂F₁₆INO₃S: C, 15.42; H, 0.32; N, 2.25. Found: C, 15.51; H, <0.3; N, 2.43.

1g: white solid; mp 71–72 °C. Yield: 98%. ¹H NMR: δ 5.36 (s, 2H). ¹⁹F NMR: δ –124.6 (m, 2F), –121.5 (s, 2F), –120.7 (s, 2F), –119.5 (s, 2F), –115.7 (s, 2F), –82.1 (m, 2F), –80.5 (t, *J* = 12.4 Hz, 2F), –67.4 (t, *J* = 12.3 Hz, 2F). MS (ESI, *m/z*, %): 530.0 [M]⁻ (100). IR (KBr): 3384, 3275, 1374, 1329, 1209, 1145, 1100, 1052, 998, 940, 882, 707, 698, 685, 656, 627, 547, 491 cm⁻¹. Anal. Calcd. for C₈H₂ClF₁₆NO₃S: C, 18.07; H, 0.38; N, 2.63. Found: C, 17.93; H, <0.3; N, 2.80.

1i: white solid. Yield: 94%. ¹H NMR (CD₃COCD₃): δ 8.12 (s, 4H). ¹⁹F NMR (CD₃COCD₃): δ –125.9 (m, 4F), –117.9 (s, 4F), –83.5 (m, 4F), –81.8 (m, 4F).

4.4. Typical procedure for the preparation of 1h

In a 100-mL three-neck flask equipped with katathermometer and magnetic stirrer, $FO_2SCF_2CF_2CCF_2CF_2SO_2F$ (5.02 g, 13.1 mmol) was added dropwise into liquid NH₃ (30 mL) within 40 min. The reaction mixture was stirred at -78 °C for 2 h. After removing excessive ammonia, the residue was acidified with diluted H₂SO₄ till pH value below 2. Then water was evaporated. The crude product was extracted with acetone and purified by column chromatography on silica gel using dichloromethane/acetonitrile (4:1) as the eluent. 4.82 g of pure **1h** (12.8 mmol, 97%) was obtained as a white solid. mp 125–126 °C. ¹H NMR (CD₃COCD₃): δ 8.03 (s, 4H). ¹⁹F NMR (CD₃COCD₃): δ –117.2 (s, 4F), –81.2 (s, 4F). MS (ESI, *m/z*, %): 375.0 [M]⁻ (100). IR (KBr): 3406, 3307, 1532, 1376, 1347, 1305, 1232, 1183, 1156, 1114, 916, 771, 655, 637, 609, 565, 503, 481, 419 cm⁻¹. Anal. Calcd. for C₄H₄F₈N₂O₅S₂: C, 12.77; H, 1.07; N, 7.45. Found: C, 12.94; H, 0.79; N, 7.31.

4.5. Typical procedure for the preparation of 2aa-ah, 2ak, 2da-di, 2ha-hd and 2ia-id

In a 15 mL round bottom sealed tube equipped with a magnetic stirrer, **1d** (0.220 g, 0.740 mmol) and 1-iodobutane (0.278 g, 1.51 mmol) were dissolved in THF (7 mL). K₂CO₃ (0.196 g, 1.42 mmol) was then added. After being heated at 90 °C for 2 days, the reaction mixture was cooled, concentrated and acidified with diluted H₂SO₄ till pH < 2. The resulting emulsion was extracted with ethyl ether (30 mL), washed by water (3 × 15 mL) and dried over anhydrous Na₂SO₄. The crude product was purified by column chromatography to give pure **2de** (0.112 g, 0.317 mmol, 43%) and **2df** (0.137 g, 0.335 mmol, 45%) using petroleum ether/ethyl acetate (20:1) as the eluent.

2de: colorless liquid. Yield: 43%. ¹H NMR: δ 5.87 (tt, *J* = 52.6 Hz, *J* = 3.2 Hz, 1H), 4.82 (s, 1H), 3.33 (q, *J* = 6.8 Hz, 2H), 1.58 (m, 2H), 1.39 (m, 2H), 0.95 (t, *J* = 7.1 Hz, 3H). ¹⁹F NMR: δ -137.2 (dt, *J* = 52.6 Hz, *J* = 5.2 Hz, 2F), -116.1 (s, 2F), -88.3 (m, 2F), -81.5 (m, 2F). ¹³C NMR: δ 44.6, 32.3, 19.4, 13.4. MS (EI, *m/z*, %): 310 (100), 136 (36.0), 128 (29.8), 119 (44.3), 101 (42.7), 78 (19.9), 57 (38.3), 56 (30.5). IR (KBr): 3313, 3133, 2968, 2881, 1430, 1398, 1330, 1285, 1200, 1143, 1078, 857, 749, 612, 583 cm⁻¹. Anal. Calcd. for C₈H₁₁F₈NO₃S: C, 27.20; H, 3.14; N, 3.97. Found: C, 27.46; H, 3.19; N, 3.88.

2df: colorless liquid. Yield: 45%. ¹H NMR: δ 5.86 (tt, *J* = 52.5 Hz, *J* = 3.2 Hz, 1H), 3.37 (d, *J* = 37.0 Hz, 4H), 1.61 (s, 4H), 1.34 (m, 4H), 0.95 (t, *J* = 6.8 Hz, 6H). ¹⁹F NMR: δ –137.3 (dt, *J* = 52.5 Hz, *J* = 5.1 Hz, 2F), –115.9 (s, 2F), –88.5 (m, 2F), –81.7 (m, 2F). ¹³C NMR: δ 48.3, 30.5, 19.7, 13.6. MS (EI, *m/z*, %): 366 (17.8), 324 (100), 310 (26.8), 192 (16.1), 119 (20.1), 101 (17.5), 57 (38.6), 43 (14.6). IR (KBr): 2964, 2939, 2879, 1469, 1390, 1326, 1283, 1143, 1034, 925, 857, 748, 582 cm⁻¹. Anal. Calcd. for C₁₂H₁₉F₈NO₃S: C, 35.21; H, 4.68; N, 3.42. Found: C, 35.18; H, 4.80; N, 3.50.

2aa: colorless liquid. Yield: 89%. ¹H NMR: δ 3.07 (s, 6H). ¹⁹F NMR: δ –75.1 (s, 3F).

2ab: colorless liquid. Yield: 90%. ¹H NMR: δ 3.46 (q, *J* = 6.8 Hz, 4H), 1.26 (t, *J* = 6.8 Hz, 6H). ¹⁹F NMR: δ –77.0 (s, 3F).

2ac: white solid. Yield: 16%. ¹H NMR: δ 4.79 (s, 1H), 3.28 (q, *J* = 6.4 Hz, 2H), 1.65 (m, 2H), 0.99 (t, *J* = 6.4 Hz, 3H). ¹⁹F NMR: δ –78.0 (s, 3F).

2ad: colorless liquid. Yield: 46%. ¹H NMR: δ 3.31 (t, *J* = 7.3 Hz, 4H), 1.67 (m, 4H), 0.94 (t, *J* = 7.3 Hz, 6H). ¹⁹F NMR: δ -76.4 (s, 3F). ¹³C NMR: δ 120.1 (q, *J* = 323.7 Hz, CF₃), 50.0, 21.7, 10.9. MS (EI, *m/z*, %): 233 (12.3), 204 (100), 164 (12.8), 162 (38.2), 69 (8.4), 43 (66.8), 42 (11.4), 41 (13.4). IR (KBr): 2973, 2942, 2882, 1715, 1506, 1471, 1390, 1226, 1200, 1133, 1005, 874, 801, 741, 599, 511 cm⁻¹. Anal. Calcd. for C₇H₁₄F₃NO₂S: C, 36.04; H, 6.05; N, 6.00. Found: C, 35.99; H, 5.97; N, 5.75.

2ae: colorless liquid. ¹H NMR: δ 4.77 (s, 1H), 3.31 (q, *J* = 6.8 Hz, 2H), 1.60 (m, 2H), 1.39 (m, 2H), 0.95 (t, *J* = 7.3 Hz, 3H). ¹⁹F NMR: δ –78.0 (s, 3F).

2af: colorless liquid. Yield: 77%. ¹H NMR: δ 3.34 (t, *J* = 6.8 Hz, 4H), 1.61 (m, 4H), 1.34 (m, 4H), 0.95 (t, *J* = 7.3 Hz, 6H). ¹⁹F NMR: δ -76.4 (s, 3F). ¹³C NMR: δ 120.1 (q, *J* = 323.7 Hz, CF₃), 48.1, 30.4, 19.7, 13.6. MS (EI, *m/z*, %): 218 (13.6), 178 (5.3), 176 (100), 69 (2.9), 57 (21.8), 56 (6.3), 42 (4.6), 41 (9.1). IR (KBr): 2964, 2938, 2878, 1469, 1389, 1225, 1189, 1133, 1030, 926, 597, 510 cm⁻¹. Anal. Calcd. for C₉H₁₈F₃NO₂S: C, 41.37; H, 6.94; N, 5.36. Found: C, 41.45; H, 6.87; N, 5.20.

2ag: white solid. Yield: 6%. ¹H NMR: δ 7.42–7.32 (m, 5H), 5.03 (s, 1H), 4.45 (d, *J* = 5.9 Hz, 2H). ¹⁹F NMR: δ –77.8 (s, 3F).

2ah: white solid. Yield: 29%. ¹H NMR: δ 7.37–7.33 (m, 6H), 7.20–7.17 (m, 4H), 4.43 (s, 4H). ¹⁹F NMR: δ –76.1 (s, 3F).

2ak: colorless liquid. Yield: 41%. ¹H NMR: δ 3.77 (t, *J* = 3.2 Hz, 4H), 3.51 (s, 4H). ¹⁹F NMR: δ –75.6 (s, 3F).

2da: colorless liquid. Yield: 88%. ¹H NMR: δ 5.87 (tt, *J* = 52.5 Hz, *J* = 3.2 Hz, 1H), 3.09 (s, 6H). ¹⁹F NMR: δ -138.0 (dt, *J* = 52.6 Hz, *J* = 5.2 Hz, 2F), -115.9 (s, 2F), -89.1 (m, 2F), -82.7 (m, 2F). ¹³C NMR: δ 38.2. MS (EI, *m/z*, %): 119 (10.5), 110 (5.4), 108 (100), 101 (14.5), 92 (12.3), 51 (6.4), 44 (10.0), 42 (11.1). IR (KBr): 2962, 1424, 1386, 1329, 1286, 1200, 1135, 1001, 967, 857, 719, 581, 489 cm⁻¹. Anal. Calcd. for C₆H₇F₈NO₃S: C, 22.16; H, 2.17; N, 4.31. Found: C, 22.15; H, 2.22; N, 4.24.

2db: colorless liquid. Yield: 89%. ¹H NMR: δ 5.86 (tt, *J* = 52.6 Hz, *J* = 3.2 Hz, 1H), 3.48 (d, *J* = 27.0 Hz, 4H), 1.26 (t, *J* = 7.3 Hz, 6H). ¹⁹F NMR: δ –138.0 (dt, *J* = 52.6 Hz, *J* = 5.2 Hz, 2F), –117.0 (s, 2F), –89.2 (m, 2F), –82.3 (m, 2F).

2dc: colorless liquid. Yield: 37%. ¹H NMR: δ 5.87 (tt, *J* = 52.5 Hz, *J* = 3.2 Hz, 1H), 3.41 (s, 1H), 3.30 (t, *J* = 6.8 Hz, 2H), 1.64 (m, 2H), 0.98 (t, *J* = 7.8 Hz, 3H). ¹⁹F NMR: δ –137.9 (dt, *J* = 52.6 Hz, *J* = 5.2 Hz, 2F), –116.8 (s, 2F), –89.0 (m, 2F), –82.2 (m, 2F). ¹³C NMR: δ 46.5, 23.6, 10.7. MS (EI, *m/z*, %): 310 (100), 128 (27.4), 122 (76.4), 119 (47.0), 106 (18.5), 101 (47.5), 78 (21.6), 43 (43.9). IR (KBr): 3317, 2976, 2886, 1432, 1372, 1330, 1285, 1200, 1143, 1075, 986, 857, 750, 612, 584 cm⁻¹. HRMS for C₇H₉F₈NO₃S: 339.0175. Found: 339.0171.

2dd: colorless liquid. Yield: 41%. ¹H NMR: δ 5.86 (tt, *J* = 52.5 Hz, *J* = 3.2 Hz, 1H), 3.33 (d, *J* = 32.0 Hz, 4H), 1.66 (m, 4H), 0.94 (t, *J* = 7.3 Hz, 6H). ¹⁹F NMR: δ –137.9 (dt, *J* = 52.5 Hz, *J* = 5.1 Hz, 2F), –116.6 (s, 2F), –89.1 (m, 2F), –82.3 (m, 2F). ¹³C NMR: δ 50.2, 21.8, 10.8. MS (EI, *m*/*z*, %): 352 (100), 310 (25.7), 164 (44.2), 119 (24.0), 101 (15.6), 70 (15.4), 43 (59.9), 42 (15.3). IR (KBr): 2974, 2943, 2884, 1472, 1390, 1327, 1284, 1203, 1139, 1015, 976, 873, 857, 744, 583, 524 cm⁻¹. Anal. Calcd. for C₁₀H₁₅F₈NO₃S: C, 31.50; H, 3.97; N, 3.67. Found: C, 31.80; H, 4.22; N, 3.62.

2dg: colorless liquid. Yield: 88%. ¹H NMR: δ 7.32 (m, 6H), 7.18 (m, 4H), 5.87 (tt, *J* = 52.5 Hz, *J* = 3.2 Hz, 1H), 4.45 (s, 4H). ¹⁹F NMR: δ –137.9 (dt, *J* = 52.6 Hz, *J* = 5.1 Hz, 2F), –115.8 (s, 2F), –89.1 (m, 2F), –82.0 (m, 2F). ¹³C NMR: δ 133.7, 128.8, 128.4, 51.4. MS (EI, *m/z*, %): 386 (5.0), 196 (6.0), 195 (7.6), 194 (11.7), 93 (5.0), 92 (67.9), 91 (100), 65 (8.4). IR (KBr): 3035, 2360, 1508, 1458, 1389, 1326, 1200, 1144, 1059, 932, 895, 748, 698, 610, 505 cm⁻¹. Anal. Calcd. for C₁₈H₁₅F₈NO₃S: C, 45.29; H, 3.17; N, 2.93. Found: C, 45.07; H, 3.37; N, 2.96.

2dh: colorless liquid. Yield: 75%. ¹H NMR: δ 5.87 (tt, *J* = 52.6 Hz, *J* = 3.2 Hz, 1H), 5.82–5.68 (m, 2H), 5.35–5.25 (m, 4H), 3.98 (d, *J* = 5.0 Hz, 4H). ¹⁹F NMR: δ –138.0 (dt, *J* = 52.6 Hz, *J* = 5.2 Hz, 2F), –116.9 (s, 2F), –89.1 (m, 2F), –82.1 (m, 2F). ¹³C NMR: δ 131.4, 120.8, 50.0 MS (EI, *m*/*z*, %): 348 (14.5), 160 (25.6), 119 (15.4), 101 (15.6), 96 (17.7), 69 (20.4), 68 (14.4), 41 (100). IR (KBr): 3091, 2993, 2939, 1646, 1447, 1423, 1390, 1328, 1284, 1200, 1144, 1059, 992, 936, 910, 857, 778, 749, 615, 578, 504 cm⁻¹. Anal. Calcd. for C₁₀H₁₁F₈NO₃S: C, 31.84; H, 2.94; N, 3.71. Found: C, 32.04; H, 3.09; N, 3.67.

2di: colorless liquid. Yield: 71%. ¹H NMR: δ 5.87 (tt, *J* = 52.6 Hz, *J* = 3.2 Hz, 1H), 3.77 (s, 4H), 3.56 (s, 4H). ¹⁹F NMR: δ –137.8 (dt, *J* = 52.6 Hz, *J* = 5.2 Hz, 2F), -116.2 (s, 2F), -88.8 (m, 2F), -82.2 (m, 2F). ¹³C NMR: δ 66.5, 46.9. MS (EI, *m/z*, %): 119 (100), 107 (38.8), 106 (59.4), 66 (65.7), 54 (74.6), 52 (26.4), 43 (49.7), 41 (37.3). IR (KBr): 2980, 2936, 2871, 1456, 1394, 1330, 1286, 1266, 1200, 1118, 1078, 991, 958, 857, 751, 711, 699, 610, 578, 495 cm⁻¹. Anal. Calcd. for C₈H₉F₈NO₄S: C, 26.17; H, 2.47; N, 3.81. Found: C, 26.32; H, 2.46; N, 3.77.

2ha: white solid; mp 72–73 °C. Yield: 35%. ¹H NMR: δ 3.09 (s, 12H). ¹⁹F NMR: δ –115.3 (s, 4F), –82.3 (s, 4F). ¹³C NMR: δ 38.2. MS (EI, *m*/*z*, %): 110 (4.9), 109 (3.5), 108 (100), 100 (5.1), 92 (18.6), 44 (18.9), 43 (5.8), 42 (12.2). IR (KBr): 2920, 2851, 1491, 1459, 1394,

1382, 1293, 1224, 1164, 1130, 1115, 1066, 975, 751, 705, 641, 599, 539, 482 cm⁻¹. Anal. Calcd. for C₈H₁₂F₈N₂O₅S₂: C, 22.23; H, 2.80; N, 6.48. Found: C, 22.68; H, 2.90; N, 6.38.

2hb: colorless liquid. Yield: 26%. ¹H NMR: δ 3.49 (d, *J* = 30.1 Hz, 8H), 1.26 (t, *J* = 7.3 Hz, 12H). ¹⁹F NMR: δ -116.3 (s, 4F), -82.0 (s, 4F). ¹³C NMR: δ 42.9, 14.1. MS (EI, *m*/*z*, %): 473 (9.5), 136 (100), 120 (44.5), 72 (7.9), 71 (15.2), 56 (13.7), 44 (21.7), 42 (12.5). IR (KBr): 2988, 2947, 2887, 1509, 1459, 1389, 1334, 1299, 1214, 1138, 1023, 945, 792, 695, 585, 504 cm⁻¹. Anal. Calcd. for C₁₂H₂₀F₈N₂O₅S₂: C, 29.51; H, 4.13; N, 5.74. Found: C, 29.93; H, 4.20; N, 5.72.

2hc: light yellow liquid. Yield: 31%. ¹H NMR: δ 5.83–5.69 (m, 4H), 5.34–5.24 (m, 8H), 3.98 (d, *J* = 5.0 Hz, 8H). ¹⁹F NMR: δ –116.2 (s, 4F), –81.8 (s, 4F). ¹³C NMR: δ 131.5, 120.7, 50.0 MS (EI, *m/z*, %): 160 (28.3), 96 (83.4), 94 (23.1), 81 (15.3), 69 (36.7), 68 (15.8), 56 (15.1), 41 (100). IR (KBr): 3091, 2997, 2943, 2875, 1647, 1448, 1423, 1388, 1338, 1299, 1226, 1200, 1144, 992, 934, 910, 782, 737, 668, 603, 501 cm⁻¹. HRMS for C₁₆H₂₀F₈N₂O₅S₂: 536.0686. Found: 536.0682.

2hd: white solid; mp 112–113 °C. Yield: 28%. ¹H NMR: δ 7.32–7.29 (m, 12H), 7.17 (m, 8H), 4.46 (s, 8H). ¹⁹F NMR: δ –115.0 (s, 4F), –81.6 (s, 4F). ¹³C NMR: δ 133.8, 128.8, 128.7, 128.4, 51.4. MS (MALDI, *m/z*, %): 759.4 [M+Na], 775.3 [M+K]. IR (KBr): 3036, 1498, 1459, 1376, 1339, 1303, 1201, 1144, 1092, 1062, 938, 925, 906, 889, 796, 767, 742, 726, 693, 603, 509 cm⁻¹. Anal. Calcd. for C₃₂H₂₈F₈N₂O₅S₂: C, 52.17; H, 3.83; N, 3.80. Found: C, 52.36; H, 4.03; N, 3.57.

2ia: white solid; mp 81–83 °C. Yield: 30%. ¹H NMR: δ 3.10 (s, 12H). ¹⁹F NMR: δ –126.2 (m, 4F), –115.8 (s, 4F), –83.6 (m, 4F), –82.5 (s, 4F). ¹³C NMR: δ 38.2. MS (EI, *m/z*, %): 119 (3.7), 110 (4.9), 109 (3.6), 108 (100), 100 (3.4), 92 (13.9), 44 (8.7), 42 (5.3). IR (KBr): 1379, 1336, 1287, 1213, 1150, 1129, 1001, 969, 881, 701, 658, 573, 485 cm⁻¹. Anal. Calcd. for C₁₂H₁₂F₁₆N₂O₆S₂: C, 22.23; H, 1.87; N, 4.32. Found: C, 22.34; H, 1.92; N, 4.33.

2ib: colorless liquid. Yield: 27%. ¹H NMR: δ 3.48 (d, *J* = 25.1 Hz, 8H), 1.26 (t, *J* = 7.3 Hz, 12H). ¹⁹F NMR: δ -126.2 (m, 4F), -116.7 (s, 4F), -83.6 (m, 4F), -82.2 (s, 4F). ¹³C NMR: δ 42.8, 14.1. MS (EI, *m/z*, %): 689 (7.7), 137 (5.9), 136 (100), 120 (19.0), 119 (8.1), 71 (8.3), 56 (7.1), 44 (10.0). IR (KBr): 2987, 1471, 1390, 1314, 1288, 1214, 1148, 1024, 945, 791, 696, 590, 506 cm⁻¹. HRMS for C₁₆H₂₀F₁₆N₂O₆S₂: 704.0507. Found: 704.0503.

2ic: colorless liquid. Yield: 31%. ¹H NMR: δ 5.82–5.69 (m, 4H), 5.35–5.25 (m, 8H), 3.98 (d, *J* = 5.9 Hz, 8H). ¹⁹F NMR: δ –126.1 (m, 4F), –116.6 (s, 4F), –83.5 (m, 4F), –82.0 (s, 4F). ¹³C NMR: δ 131.4, 120.8, 50.0. MS (EI, *m*/*z*, %): 160 (60.6), 144 (19.3), 96 (75.3), 94 (21.7), 81 (16.0), 69 (33.6), 68 (17.4), 41 (100). IR (KBr): 3091, 2992, 2937, 1645, 1446, 1423, 1395, 1333, 1288, 1217, 1148, 1058, 991, 935, 910, 892, 766, 720, 614, 573, 504 cm⁻¹. HRMS for C₂₀H₂₀F₁₆N₂O₆S₂: 752.0507. Found: 752.0511.

2id: white solid; mp 135–136 °C. Yield: 86%. ¹H NMR: δ 7.32–7.30 (m, 12H), 7.17 (m, 8H), 4.45 (s, 8H). ¹⁹F NMR: δ –126.1 (m, 4F), –115.7 (s, 4F), –83.6 (m, 4F), –81.9 (s, 4F). ¹³C NMR: δ 133.7, 128.8, 128.7, 128.4, 51.4. MS (EI, *m/z*, %): 260 (5.0), 196 (24.0), 195 (19.9), 194 (14.7), 93 (4.8), 92 (63.4), 91 (100), 65 (4.5). IR (KBr): 3034, 1497, 1458, 1379, 1363, 1327, 1289, 1217, 1172, 1141, 1062, 1007, 936, 923, 906, 889, 744, 708, 693, 606, 690, 533, 499 cm⁻¹. Anal. Calcd. for C₃₆H₂₈F₁₆N₂O₆S₂: C, 45.38; H, 2.96; N, 2.94. Found: C, 45.39; H, 3.28; N, 2.74.

4.6. Typical procedure for the preparation of 2ai, 2aj and 2al

1a (0.152 g, 1.02 mmol) and 2-(2-hydroxyethoxy)ethyl 4methylbenzenesulfonate (0.580 g, 2.23 mmol) were dissolved in THF (10 mL) and placed in a 25 mL round bottom flask which was equipped with a magnetic stirrer. K_2CO_3 (0.286 g, 2.10 mmol) was then added. After being refluxed for 2 days, the reaction mixture was diluted by Et_2O (30 mL) and filtered. The solvent was evaporated and the crude product was purified by column chromatography to give 0.232 g of pure **2ai** (colorless liquid, 0.714 mmol, 71%) using dichloromethane/methanol (20:1) as the eluent. ¹H NMR: δ 3.75–3.59 (m, 16H), 2.89 (s, 2H). ¹⁹F NMR: δ –75.7 (s, 3F). ¹³C NMR: δ 120.0 (q, *J* = 323.8 Hz, CF₃), 72.6, 69.7, 61.6, 49.6. MS (EI, *m*/*z*, %): 190 (15.8), 176 (16.2), 118 (31.4), 88 (28.5), 75 (17.9), 56 (25.5), 45 (100), 44 (17.7). IR (KBr): 3400, 2938, 2877, 1385, 1226, 1191, 1127, 1066, 1004, 701, 594 cm⁻¹. Anal. Calcd. for C₉H₁₈F₃NO₆S: C, 33.23; H, 5.58; N, 4.31. Found: C, 32.95; H, 5.46; N, 4.00.

2aj: colorless liquid. Yield: 68%. ¹H NMR: δ 3.72–3.65 (m, 20H), 3.58 (m, 4H), 3.16 (s, 2H). ¹⁹F NMR: δ –75.9 (s, 3F). ¹³C NMR: δ 119.9 (q, *J* = 323.1 Hz, CF₃), 72.7, 70.3, 70.1, 69.9, 61.5, 49.1. MS (EI, *m/z*, %): 220 (15.1), 176 (17.4), 174 (28.2), 162 (17.7), 89 (58.4), 87 (25.6), 56 (20.1), 45 (100). IR (KBr): 3414, 2877, 1456, 1386, 1355, 1225, 1193, 1126, 1070, 1004, 701, 595 cm⁻¹. Anal. Calcd. for C₁₃H₂₆F₃NO₈S: C, 37.77; H, 6.34; N, 3.39. Found: C, 37.41; H, 6.60; N, 3.24.

2al: light yellow liquid. Yield: 9%. ¹H NMR: δ 3.70 (m, 16H), 3.62 (s, 8H). ¹⁹F NMR: δ -76.0 (s, 6F). ¹³C NMR: δ 119.9 (q, *J* = 323.9 Hz, CF₃), 70.5, 70.1, 49.3. MS (EI, *m*/*z*, %): 393 (82.3), 220 (17.6), 100 (39.1), 70 (20.3), 69 (28.1), 56 (100), 45 (18.6), 42 (24.2). IR (KBr): 2876, 1456, 1386, 1359, 1226, 1190, 1125, 1002, 769, 730, 701, 595 cm⁻¹. HRMS for C₁₄H₂₄F₆N₂O₈S₂: 526.0878. Found: 526.0871.

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